

# CK NAC (IFCC)

## INTENDED USE

Bioline CKNAC for the quantitative determination of Creatine Kinase in human serum.

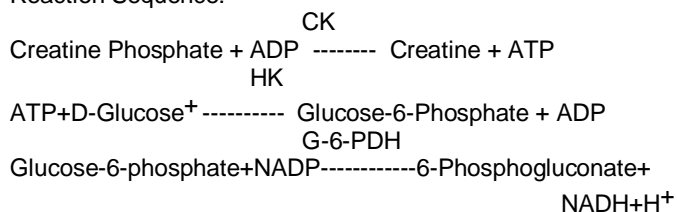
## CLINICAL SIGNIFICANCE

Creatine Kinase (CK) plays an important role in the energy-storing mechanism of tissue by catalyzing the reversible reaction between creatine and ATP to form creatine phosphate and ADP. CK is distributed in various organs; the highest activities (in decreasing order) are skeletal muscle, heart, and brain.<sup>1</sup> Thus, determination of CK is an aid in diagnosing muscular dystrophy and other diseases of the skeletal muscles, myocardial infarction, hypothyroidism, renal diseases, and/or dysfunction.<sup>2</sup>

## METHOD AND PRINCIPLE

The early procedure for determining CK was based on the rate of ATP formation.<sup>3</sup> A modified method was described by Nielson by adding a sulfhydryl compound and AMP to assure maximum CK activity and inhibit adenylate kinase activity.<sup>4</sup> Optimized conditions for measuring CK were published by Szasz in 1976 as well as by the Scandinavian committee on enzyme. The above procedure was modified again in 1979 to include EDTA. The present reagent is a modification of the above revision.

Reaction Sequence:



CK catalyzes the conversion of creatine phosphate and ADP to creatine and ATP. The ATP and glucose are converted to ADP and glucose-6-phosphate by hexokinase (HK). Glucose-6-phosphate dehydrogenase (G-6-PDH) oxidizes the D-glucose-6-phosphate and reduces the nicotinamide adenine dinucleotide (NAD). The rate of NADH formation, measured at 340nm, is directly proportional to serum CK activity.

## REAGENT COMPOSITION

When reconstituted as directed, the reagent for CK contains the following:

D-Glucose	20mM
Magnesium <sup>++</sup>	10mM
Adenosine-5'-Monophosphate (AMP)	50mM
N-Acetylcysteine (NAC)	20mM
Creatine Phosphate	30mM
Adenosine-5'-Diphosphate (ADP)	2mM
Oxidized Nicotinamide Adenine Dinucleotide Phosphate	2mM
Glucose-6-Phosphate Dehydrogenase (E.C.1.1.1.49, G-6-PDH)	3,000UL
Hexokinase (E.C.2.7.1.1, HK)	3,000UL
EDTA	2mM
Buffer	100mM

## WARNINGS AND PRECAUTIONS

1. For *in vitro* diagnostic use.
2. Exercise the normal precautions required for the handling of all laboratory reagents. Pipetting by mouth is not recommended for any laboratory reagent.

## REAGENT PREPARATION

Reagent R1 and R2 are ready to use. To prepare working reagent mix R1 and R2 in the ratio of 4:1.

## STORAGE AND STABILITY OF REAGENT.

The reagent should be stored at 2-8°C. The reagent may be used until the expiration date indicated on the package label. After reconstitution, the reagent is stable for twenty-four (24) hours at room temperature or 14 days refrigerated (2-8°C).

## REAGENT DETERIORATION

1. Physical Appearance  
If reagent appears turbid deterioration may have occurred and the product should be discarded.
2. Blank Absorbance  
If the reconstituted CK REAGENT without added sample has an absorbance greater than 0.700 at 340 nm versus reagent grade water, the reagent is considered to be unsatisfactory for use and should be discarded.
3. Control Assays  
Failure to obtain accurate results in the assay of control materials may indicate reagent deterioration.
4. We cannot guarantee the stability of reagents which have been:
  - a. Transferred from their original containers.
  - b. Improperly stored prior to or during use
  - c. Contaminated during use

## SPECIMEN COLLECTION AND STABILITY

Collect whole blood by non-traumatic venipuncture and allow to clot. Centrifuge and remove serum immediately. Serum is reportedly stable for four (4) hours at room temperature, 8-12 hours at 4°C, and 2-3 days when frozen.

Hemolyzed specimens should not be used because of side reactions that may occur due to adenylate kinase, adenosine triphosphate, and glucose-6-phosphate dehydrogenase liberated from red cells.

## INTERFERENCES

Certain drugs and medications may affect the activity of CK, see Young et al.<sup>9</sup>

## MANUAL PROCEDURE

Wavelength 340nm  
Temperature 37°C

	Test
Reagent R1	800µL
Reagent R2	200µL
Sample	40µL

Mix and after 3 min of incubation measure the optical density during next 90 sec.

1. Prepare working reagent according to instructions.
2. Pipette 1.0ml of reagent into appropriate tubes and pre-warm at 37°C.
3. Zero spectrophotometer with distilled water at 340nm.
4. Add 0.040ml (40µl) of sample to the reagent, mix, and incubate at 37°C for 3 minutes.
5. After 3 minutes, read and record the absorbance. Return the tubes to 37°C. Repeat readings every 30 sec for the next 90 sec.
6. Calculate the average absorbance difference per minute ( $\Delta$ Abs./min.)
7. The Abs./min. multiplied by the factor 4127 (see Calculations) will yield results in IU/L.
8. Samples with values above 2000 IU/L should be diluted 1:1 with saline, re-assayed, and the results multiplied by two (2).

## CALCULATIONS

IU/L =  $\Delta$ A Abs./min. x 4127

## LIMITATIONS

1. Some inhibitors of CK activity<sup>10</sup>
  - a. Excessive Mg<sup>++</sup>, Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>
  - b. Most heavy earth metals, i.e. Zn<sup>++</sup>, Cu<sup>++</sup>, Mn<sup>++</sup>
  - c. Iodoacetate, and other sulfhydryl binding agents
  - d. Excess ADP, citrate, fluoride, L-thyroxine
  - e. Excess uric acid
2. This procedure measures total CK activity irrespective of its tissue or organ of origin.
3. Lower than expected CK values have been reported in samples having high alkaline phosphatase activity.

## QUALITY CONTROL

Use control sera with known normal and abnormal values to monitor the integrity of the reaction. Values should be those acceptable for this method and temperature.

## EXPECTED VALUES

25- 192 IU/L (37°C)

10- 109 IU/L (30°C)

It is strongly recommended that each laboratory establish its own normal range.

## PERFORMANCE CHARACTERISTICS

1. Linearity: 2000 IU/L
2. Comparison: Studies done between this procedure and a similar procedure yield a correlation coefficient of 0.991 with a regression equation of  $Y = 1.01X - 0.29$ .
3. Precision studies:

Mean (mg/dl)	S.D.	Within Run	C.V.%
111	1.6		1.5
373.5	12.4		3.3

Mean (mg/dl)	S.D.	Run to Run	C.V.%
110.9	4.3		3.9
367.4	10.3		2.8

<b>Mode</b>	<b>Kinetic</b>
<b>Wavelength (Filter)</b>	<b>340nm</b>
<b>Reaction Direction</b>	<b>Increasing</b>
<b>Sample Vol.</b>	<b>40µl</b>
<b>Reagent Vol.</b>	<b>1000 µl</b>
<b>Delay Time / Lag Time</b>	<b>180 Sec</b>
<b>Measuring Time</b>	<b>90 Sec</b>
<b>Reagent Blank Abs.(Max)</b>	<b>NMT 0.700</b>
<b>Calibration Method</b>	<b>Fix factor</b>
<b>Factor</b>	<b>4127</b>
<b>Linearity</b>	<b>2000 IU/L</b>
<b>Decimal Places</b>	<b>1</b>
<b>Temp.</b>	<b>37°C</b>
<b>Unit</b>	<b>IU/L</b>
<b>Ref. Low (Male / Female)</b>	<b>25 IU/L</b>
<b>Ref. High (Male / Female)</b>	<b>192 IU/L</b>

## REFERENCES

1. Faulker, W.R. and Meites, S.: *Selected Method of Clinical Chemistry*. vol 9, p. 185 (1982).
2. Rosalki, S.B.: *J. Lab. Clin. Med.* 69:696 (1967).
3. Oliver, I.T.: *Biochem. J.* 61:116 (1955).